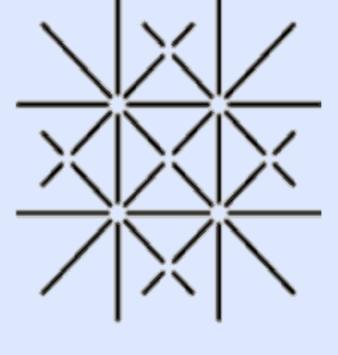
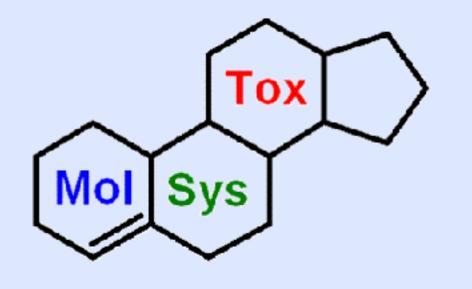
Biochemical and Molecular Modeling Analyses Explain the Functional Loss of 17β-HSD3 Mutant G133R in Three Tunisian Patients



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Introduction

17β-Hydroxysteroid dehydrogenase type 3 (17β-HSD3) catalyzes the conversion of $\Delta 4$ -androstene-3,17-dione to testosterone in testicular Leydig cells and has a key role in male sexual development¹. Mutations in the HSD17B3 gene can result in reduced enzyme activity and decreased testosterone synthesis, leading to a rare autosomal recessive disease named 46, XY disorder of sex development (46, XY DSD)².

Patients with 46, XY DSD show undervirilization of external genitalia, which often appear female³. They are usually raised as females until a virilization occurs at puberty due to extra testicular testosterone synthesis⁴ by the enzyme 17β-HSD5, which is not expressed in early development.

We characterized three Tunisian patients from non-consanguineous families with 46, XY DSD and investigated 17β-HSD3 deficiency.

Patients Clinical History

Genomic DNA from patient 1 was directly analyzed for mutations in the HSD17B3 gene by DNA sequencing because of a familial history, recording a paternal cousin with HSD17B3 deficiency. For the patients P2 and P3 a human chorionic gonadotropin stimulation test was performed, due to signs of virilization observed at puberty and the absence of a complete hormonal profile. The results revealed 17β-HSD3 deficiency.

	Patient 1 (P1)	Patient 2 (P2)	Patient 3 (P3)		
Age (years)	7	14			
Height cm	155	-	-	Primary Amenorrhea	$P \circ P \circ$
Weight Kg	45	-	-	and signs of virilisation	
Tanner stage	P1B1	P4B1	P5B1		$Q \cdot Q = Q$
Testosterone ng/ml	0.8	3	4		
LH, IU/L	12.5	-	_	2	
FSH, IU/L	4.7	40	42	4 5	<u> </u>
Caryotype analysis	46, XY	46, XY	46, XY		P2 P3

References values: LH: 1.24-8.62 mUI/ml; FSH: 1.27-19.26 mUI/ml; T: 1.75 - 7.61 ng/mL

Genetic Analysis

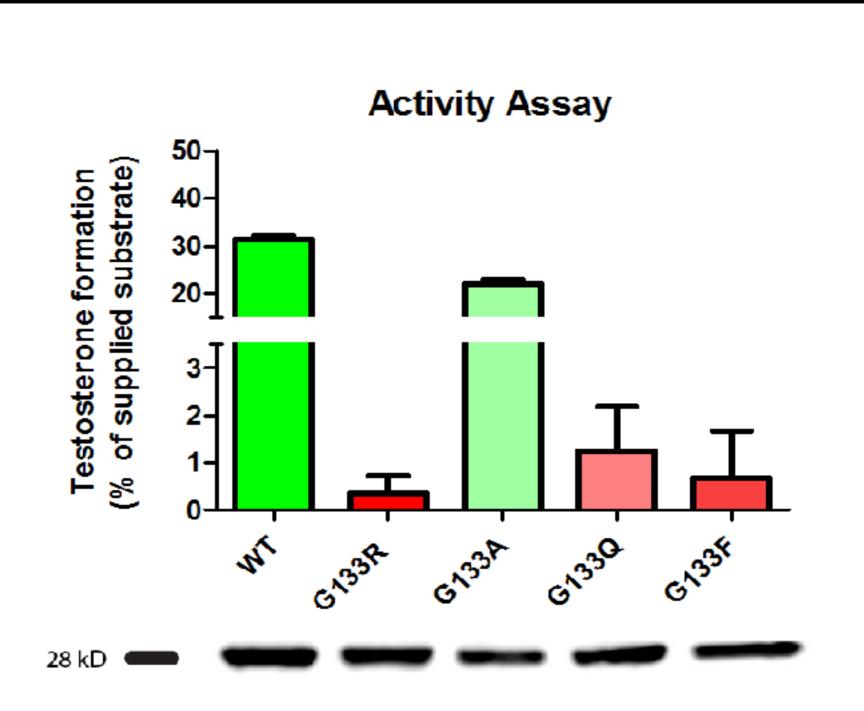
Genetic analysis of the HSD17B3 gene revealed two compound heterozygous mutations, i.e. a novel missense mutation (G133R) and a premature stop codon (C206X).

	Glycine	Arginine
17bHSD1	68	RDSKSVAAARERVT-EGRVDVLVCNAG-LGLLGPL-EALGE
17bHSD2	140	TKPVQIKDAYSKVA-AMLQDRGLWAVINNAGVLGFPTDG-ELLLM
17bHSD3	107	FTKDDIYEHIKEKL-AGLEIGILVNNVGMLPNLLPS-HFLNA
17bHSD4	71	ANYDSVEEGEKVVKTALDAFGRIDVVVNNAGILRDR-SFARI
17bHSD6	85	TKMESIAAATQWVK-EHVGDRGLWGLVNNAGILTPITLC-EWLNT
17bHSD7	65	SNLQSVFRASKELKQRFQRLDCIYLNAGIMPNPQLNIKALFF
17bHSD8	77	SEARAARCLLEQVQ-ACFSRP-PSVVVSCAGITQD-EFLLH
17bHSD9	84	TDPQSVQQAAKWVE-MHVKEAGLFGLVNNAGVAGIIGPT-PWLTR
17bHSD10	66	TSEKDVQTALALAKGKFGRVDVAVNCAGIAVASKTYNLKKGQ
17bHSD11	95	SNREDIYSSAKKVK-AEIGDVSILVNNAGVVYTS-DLFAT
17bHSD12	109	FASEDIYDKIKTGL-AGLEIGILVNNVGMS-YEYPE-YFLDV
17bHSD13	70	KVK-KEVGDVTIVVNNAGTVYPA-DLLST
17bHSD14	64	TQEDDVKTLVSETIRRFGRLDCVVNNAGHHPPPQRPE

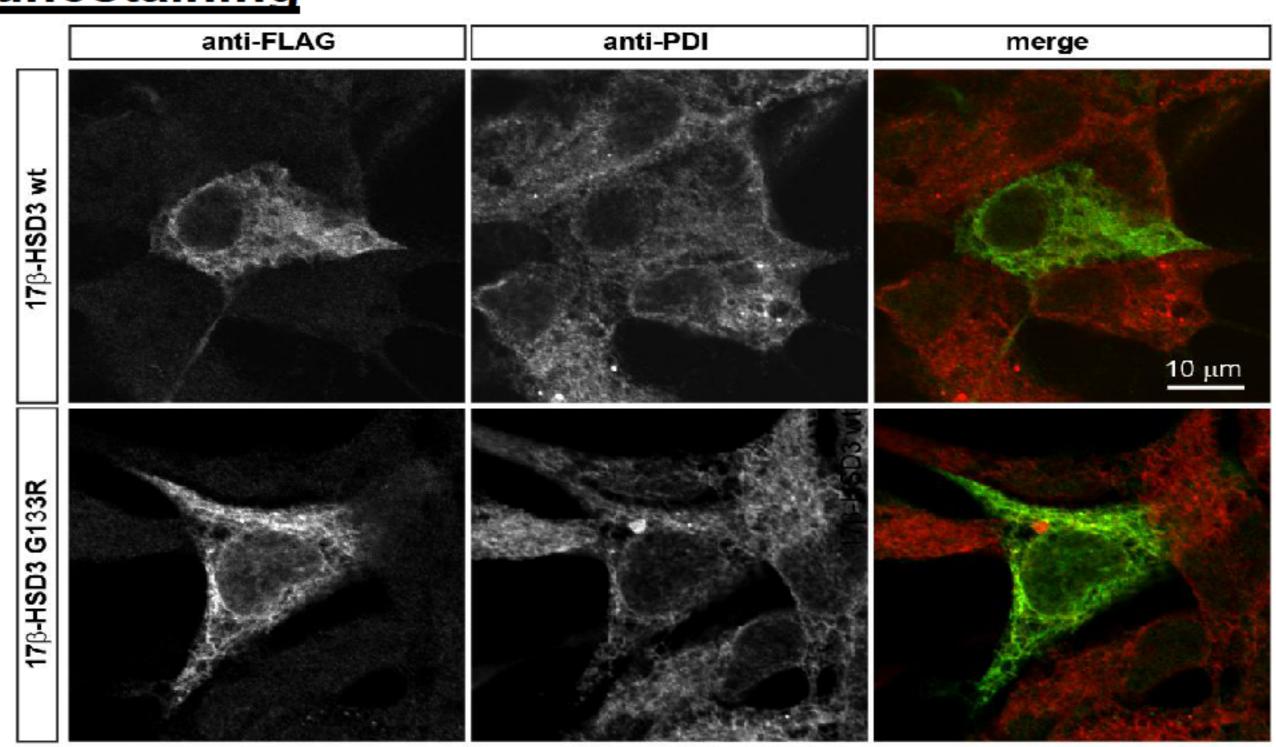
Alignment of related 17β-HSD's showed that the residue G133 and prior amino acids are highly conserved among the subfamily of short chain dehydrogenase/reductase.

Activity Assay

Using site-directed mutagenesis, an expression plasmid for 17β-HSD3 G133R was constructed. Additionally, expression plasmids substitutions of glycine 133 to alanine (G133A), to phenylalanine (G133F), and to glutamine (G133Q) were created. Wild-type and mutant enzymes were expressed in HEK-293 cells, followed by assessment of the conversion of radiolabeled Δ4-androstene-3,17-dione testosterone. Mutants G133R, G133Q and G133F were almost completely inactive, whereas G133A retained more than 80% of wild-type activity. Western blot analysis showed comparable expression of all mutants.

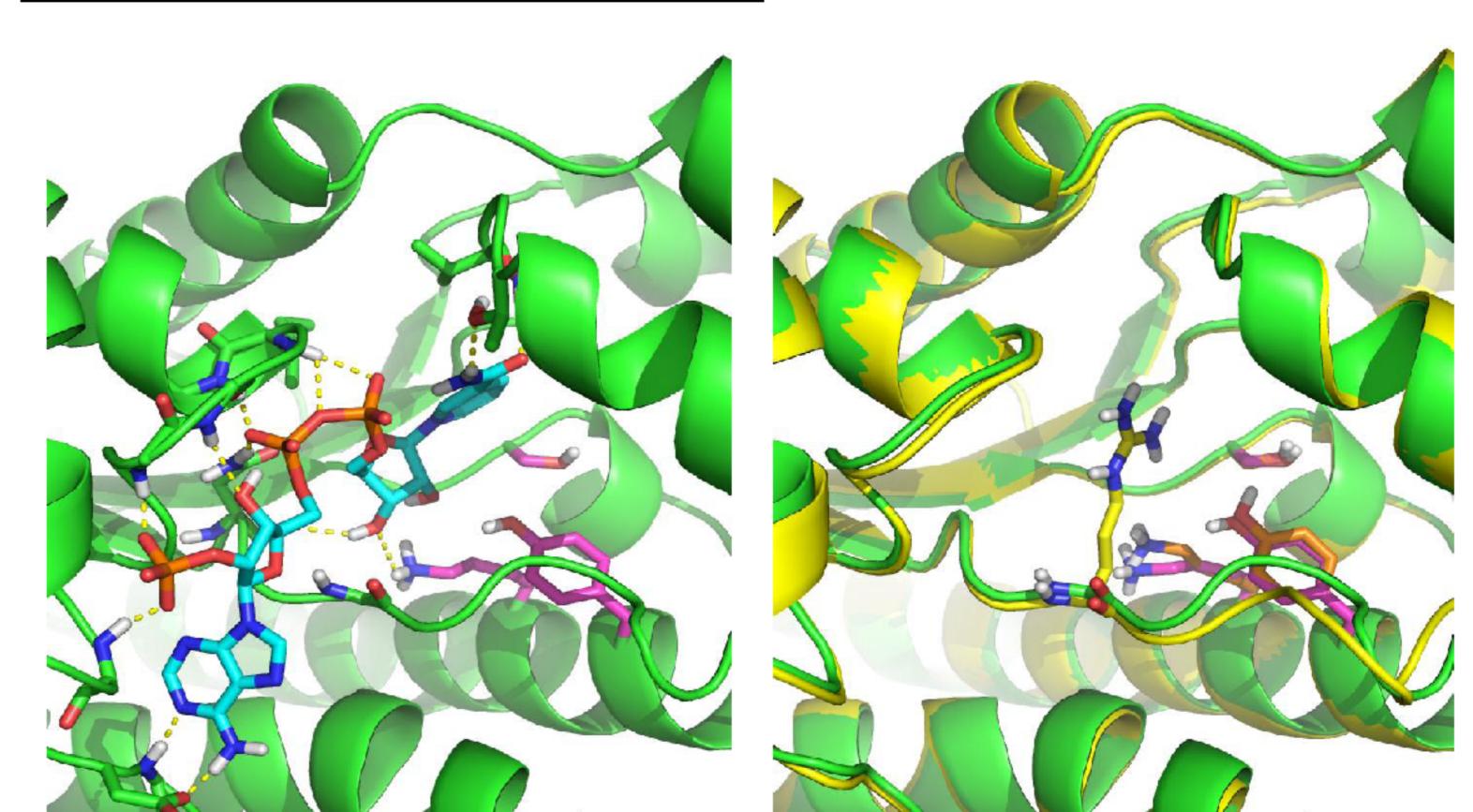


Immunostaining



Immunostaining results of 17β-HSD3-FLAG and 17β-HSD3 G133R-FLAG confirmed the expression of both enzymes at the endoplasmic reticulum membrane. No indication of dislocation nor degradation was evident.

Molecular Modeling Prediction



A homology model of 17β-HSD3 predicted that the loss of activity is due to a disruption of the cofactor binding site. While an alanine at position 133 was still tolerated, more bulky side-chains led to steric hindrance thus preventing cofactor binding.

Conclusion

We characterized three Tunisian patients from non-consanguineous families with 46, XY DSD due to 17β-HSD3 deficiency. Genetic analysis of the HSD17B3 gene revealed two compound heterozygous mutations, i.e. a novel missense mutation (G133R) and a premature stop codon (C206X). In activity measurements, mutants G133R, G133Q, and G133F were almost completely inactive, whereas G133A retained >80% of wildtype activity. A homology model of 17β-HSD3 predicted that the loss of activity is due to a disruption of the cofactor binding site. While an alanine at position 133 was still tolerated, more bulky side-chains led to steric hindrance thus preventing cofactor binding. The functional analysis and homology modeling revealed an important role of this residue in the structural arrangement of the cofactor binding pocket. The results provide an improved mechanistic understanding of the 17β-HSD3 structure-function relationship and explained the 17β-HSD3 deficiency observed in the patients.

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