Metformin regulates the differentiation of murine mesenchymal stem cells



via AMPK-independent suppression of p70s6-kinase

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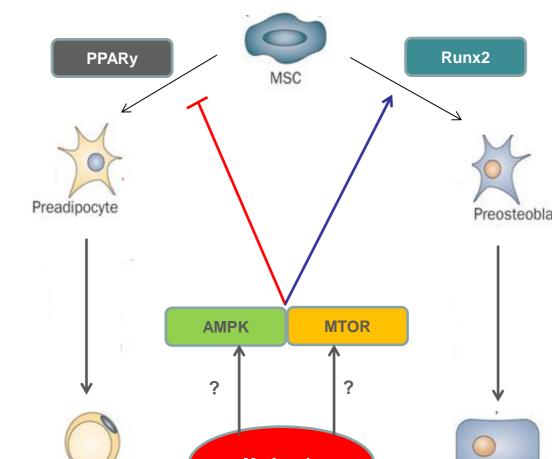
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Background and Objective

Metformin is widely used as oral anti-hyperglycaemic agent to treat Type 2 diabetes, with increasing reports of an additional, potential bone protective role. On a cellular level, mesenchymal stem cells (MSCs) have

reported to reciprocal been have differentiation into bone at the expense of fat, and vice versa. We set out to investigate the effects of metformin on the multipotent mesenchymal stem cell differentiation and the underlying molecular mechanism(s) involved.



Methods

Confluent murine MSCs (C3H10T1/2) were treated with metformin (500 μ M), a known AMPK activator (A769662;100µM) or the p70S6K inhibitor (rapamycin;10µM), in both control and adipogenic-inducing environments (using pioglitazone;10µM) for 5 days. Nuclear extracts were separated by SDS-PAGE and immunoblotted with primary antibodies to peroxisome proliferator-activated receptor gamma (PPARy;marker for adipogenesis), Runt-related transcription factor 2 (Runx2;marker for osteogenesis), phosphorylated-ACC (P-ACC(Ser79);marker for AMPK activity) and phosphorylated-p70s6k (P-p70s6k(Thr389); upstream regulator of mTOR signalling). Immunoblots were scanned using a Licor fluorescent reader. PPARy and Runx2 activities were determined using Luciferase reporter assays and adipogenesis was quantified histochemically by staining neutral lipids with Oil Red O.



50

40

30

20

10

0

control

Results

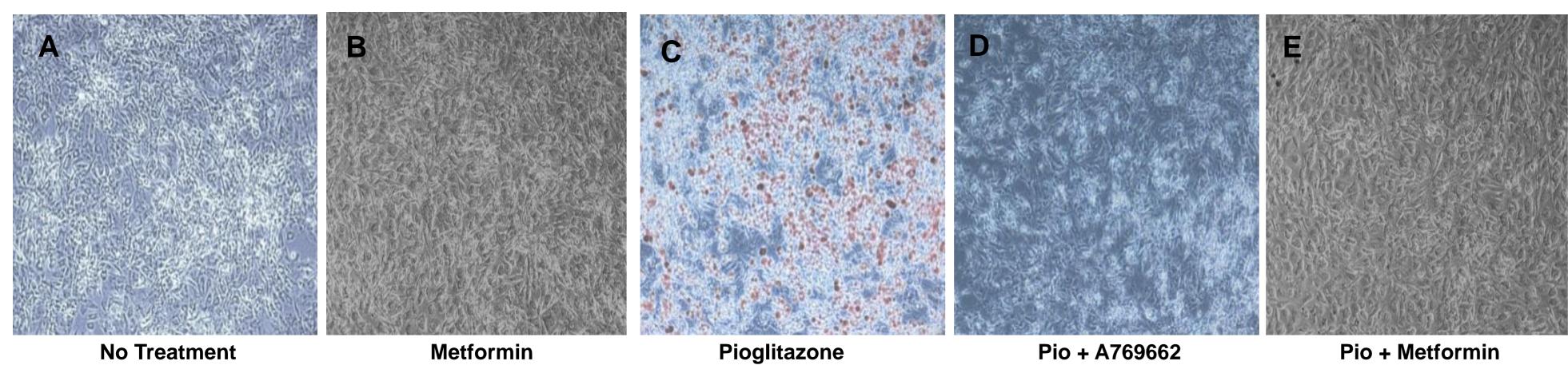
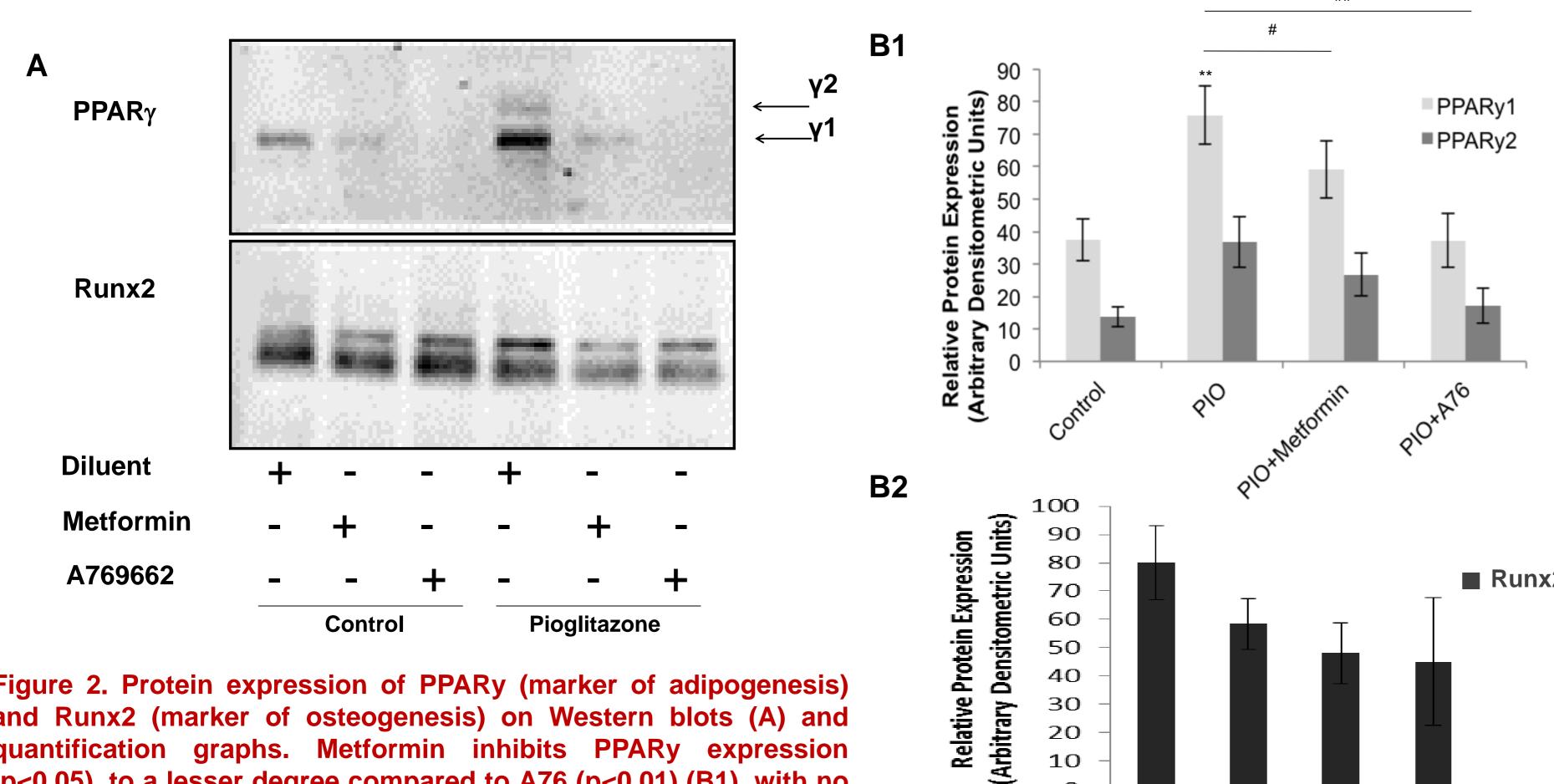
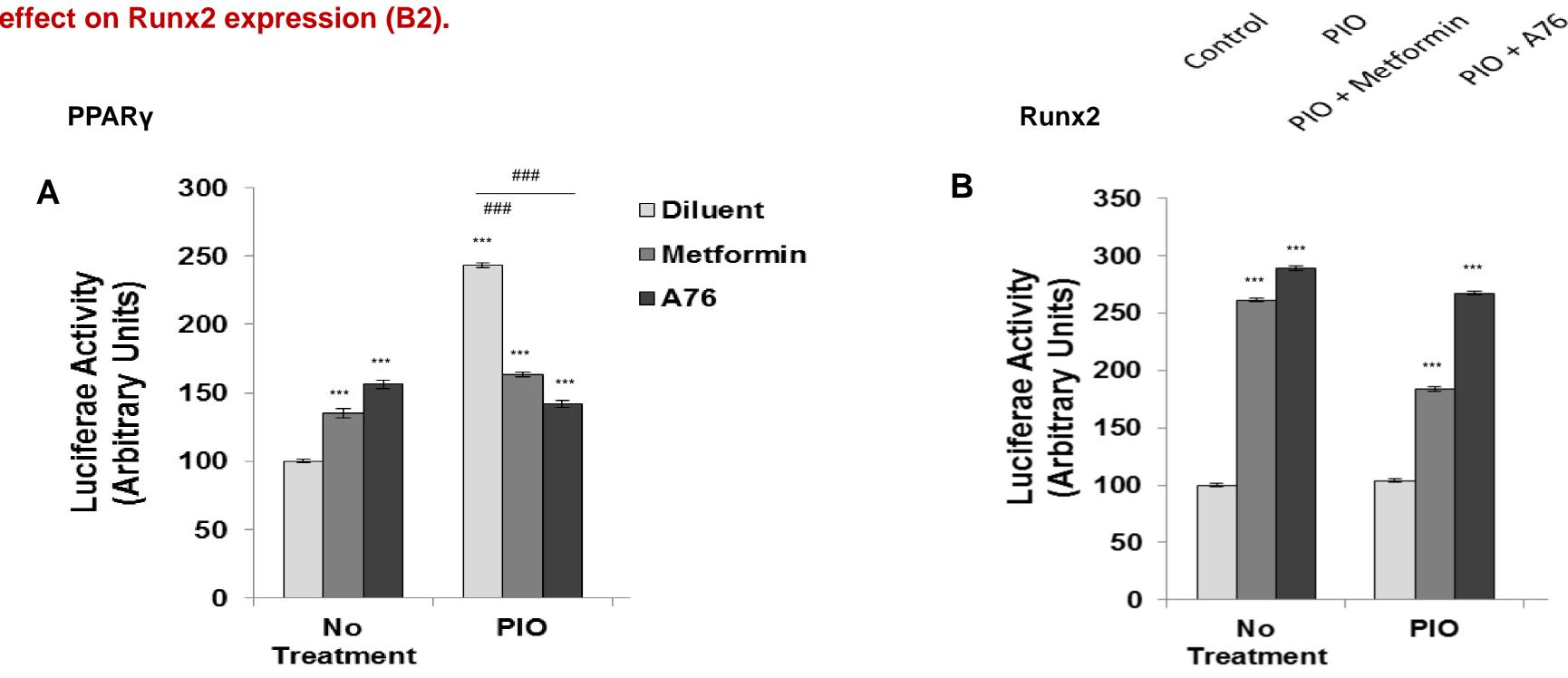


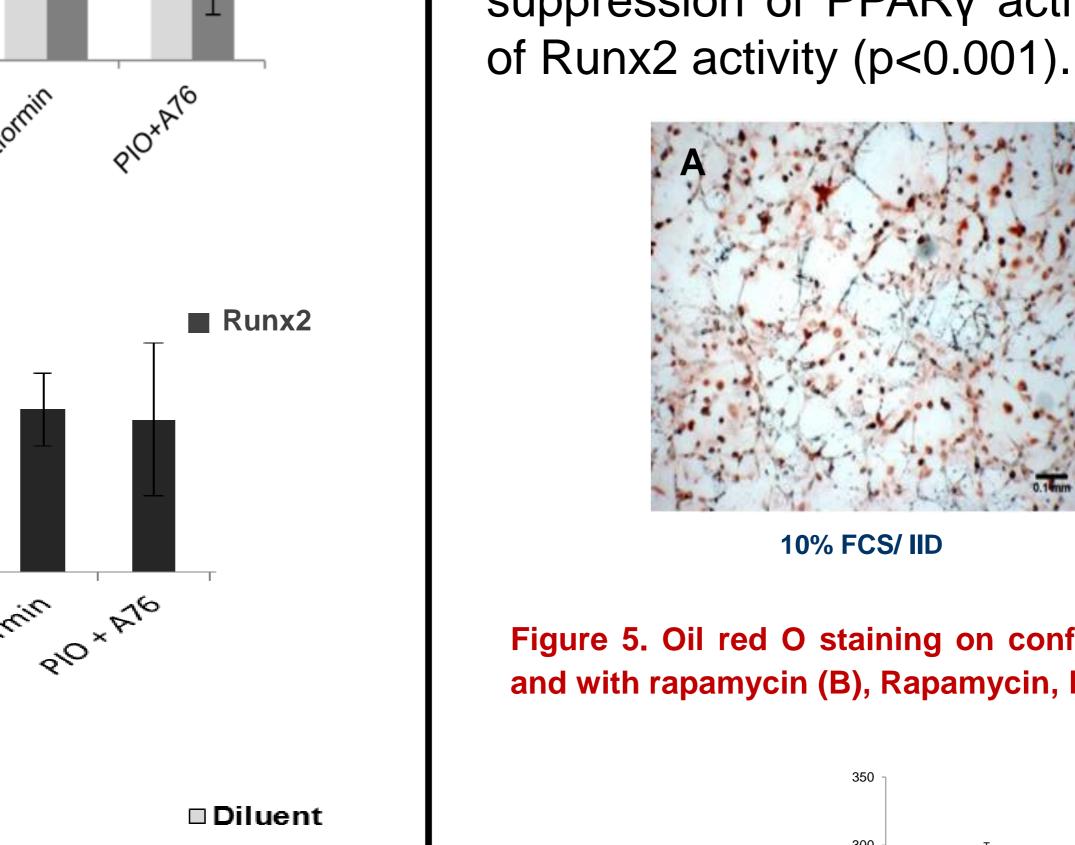
Figure 1. Oil red O staining on confluent C3H10T1/2 without treatment (A), metformin only (B), pioglitazone only (C), pioglitazone and AMPK activator A769662 (D), and pioglitazone and metformin (E). Pioglitazone exerts marked adipogenic effect whilst metformin and A769962 inhibits adipogenesis.



MSCs treated with pioglitazone demonstrated marked adipogenic phenotype staining positively with Oil Red O. In contrast, treatment with both metformin and A769662 impaired adipogenesis (Figure 1). Pioglitazone induced an (p < 0.01) increase in PPARy expression, whilst metformin and A796662 suppressed PPARy expression to basal levels, p < 0.05 and p < 0.01 respectively. Runx2 activity was significantly increased by metformin (p<0.001) and A769662 (P<0.001) but not Runx2 protein levels (Figure 2 and 3). As expected, A769662 promotes phosphorylation of ACC, but not so with metformin (Figure 4). Instead, metformin suppressed (p<0.05) the phosphorylation of p70s6k, as did A769662 (p<0.05) and rapamycin (p<0.001) (Figure 6). Luciferase reporter assays confirmed the reciprocal action of metformin on adipogenesis and osteogenesis, namely suppression of PPARy activity (p<0.001) and induction

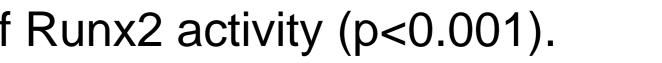
Figure 2. Protein expression of PPARy (marker of adipogenesis) and Runx2 (marker of osteogenesis) on Western blots (A) and quantification graphs. Metformin inhibits PPARy expression (p<0.05), to a lesser degree compared to A76 (p<0.01) (B1), with no effect on Runx2 expression (B2).





Metformin

■A76



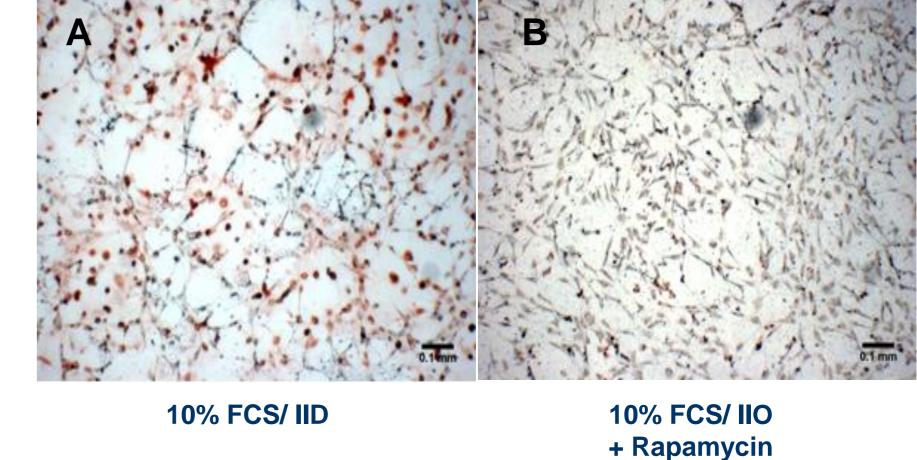


Figure 5. Oil red O staining on confluent C3H10T1/2 without treatment (A), and with rapamycin (B), Rapamycin, like metformin inhibits adipogenesis.

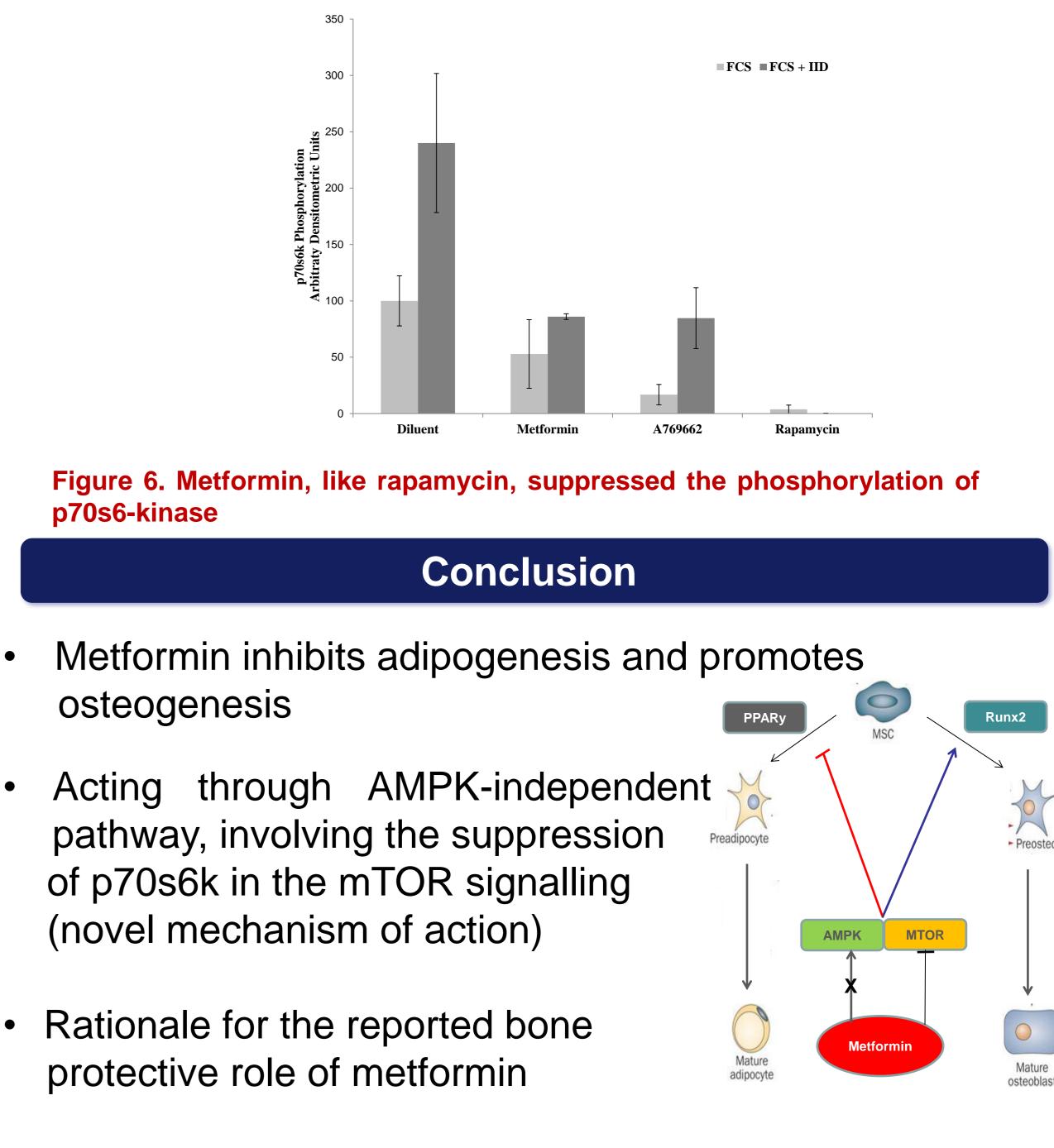


Figure 3. Luciferase assay to determine PPARy (marker of adipogenesis) and Runx2 (marker of osteogenesis) activities. Metformin reciprocally inhibits PPARy activity (p<0.001) (A) and increases Runx2 activity (p<0.001)(B).

