

## Introduction and objectives

Farnesoid X receptor (FXR) and Takeda G protein-coupled receptor 5 (TGR5) are interesting pharmacological targets for the treatment of liver and metabolic diseases. FXR-deficient mice on a high-fat diet (HFD) exhibit massive hepatic steatosis, necro-inflammation and fibrogenesis. Moreover, **pharmacological activation of TGR5 in mice promotes protective mechanisms in biliary epithelial cells, inhibits hepatic and systemic inflammation.**

The aim of this study is to investigate the effect of **FXR/TGR5 dual agonists** on nonalcoholic steatohepatitis (NASH) in a rabbit model of high fat diet (HFD)-induced MetS.

## Methods

We employed a recently established **animal model of high fat diet (HFD)-induced MetS**, characterized by insulin resistance, hypertension, atherogenic dyslipidemia, visceral adipose tissue accumulation and NASH (Filippi et al., 2009, Maneschi et al., 2013). **Subgroups of MetS rabbits were treated with increasing doses of the dual FXR/TGR5 agonist INT-767 (3, 10, 30mg/Kg, orally, daily, 5 days a week for 12 weeks).** We studied the effects of HFD and in vivo INT-767 treatments on **liver function**. Liver was studied by immunohistochemistry and RT-PCR.

## Results

- Treatment with increasing doses of the dual FXR/TGR5 agonist **INT-767** (3,10,30mg/Kg/day, 5 days a week for 12 weeks) in a rabbit model of HFD-induced MetS, characterized also by NASH, **dose-dependently reduced several MetS-associated alterations**, including hepatomegaly, insulin resistance, increase of ALT, glucose and cholesterol levels, while significantly increasing HDL levels. ALT was positively associated with all MetS parameters; however introducing all MetS factors in a multivariate analysis, only total cholesterol levels resulted positively associated with ALT level (Adj.r: 0.493, p=0.014). High macrophage M1 pro-inflammatory/M2 anti-inflammatory ratio was observed in MetS-induced NASH, which was independently associated with serum ALT levels (Adj.r: 0.322, p=0.032). **HFD-induced increase in M1/M2 ratio was reduced by INT-767 treatment and M2 macrophage markers (IL-10, TGFβ) were increased.**

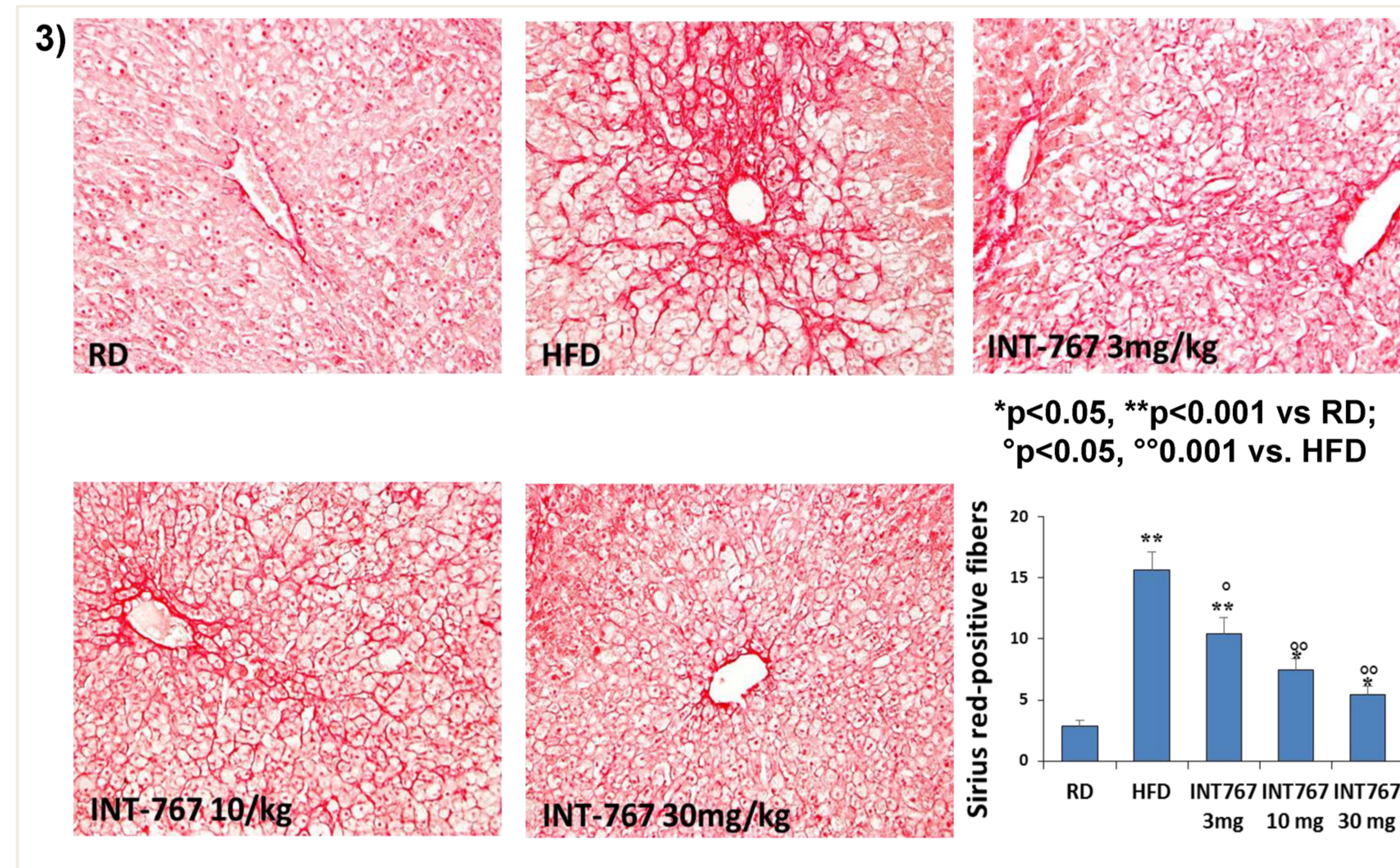
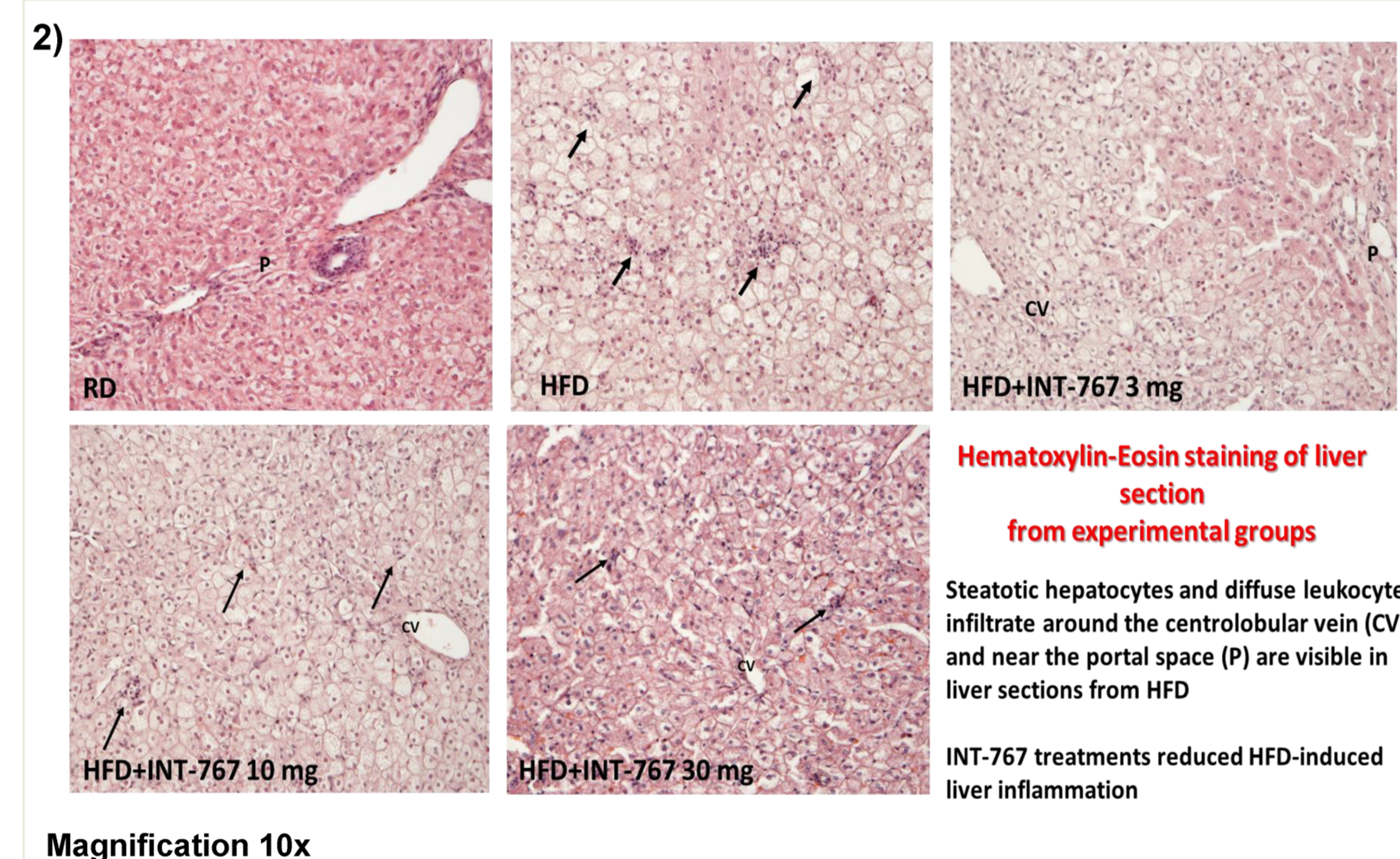
- Genes related to neutrophil apoptosis/apoptotic-neutrophil clearance (**lactoferrin, eNOS, RAGE**) and to extracellular matrix degradation (**MMP2, TIMP2**) were also **increased by INT-767** treatment. **INT-767** also **reduced** liver expression of **IL-6**, which preferentially skews the Th cell response towards a Th17-phenotype, while **increasing Foxp3** expression, a Treg cell marker (Fig. 1). Thus these data indicate that INT-767 can promote the neutrophil- and macrophage-driven resolution phase of inflammation and fibrosis regression. In addition, **INT-767 increased** genes related to hepatic fatty acid metabolism (**PPARα, AR, CD36**) and lipid droplet formation (**SNAP23, VAMP4, syntaxin5, perilipin**) therefore suggesting that INT-767 counteracts excess fatty acid mediated lipotoxicity in the liver. Genes related to insulin signaling (**IRS1, SREBP1, G6Pase**, and **PEPCK**) were also **increased by INT-767** (Fig. 1).

- Finally, immunohistochemical studies demonstrated that **INT-767** treatment significantly **reduced both HFD-induced liver inflammation** (Fig. 2) and **fibrosis**. (Fig. 3).

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\*\*p<0.01, \*\*\*p<0.0001 vs. RD; °p<0.05, °°p<0.01, °°°p<0.0001 vs. HFD

	RD	HFD	HFD+INT-767 3mg	HFD+INT-767 10mg	HFD+INT-767 30mg
M1/M2 ratio	1.1±0.2	219.9±24.5***	62.6±9.4°°	34±16°°	27.1±12°°°
M2 macrophage markers					
IL10	100±8.69	1218.28±143.19***	2779.2±560.50°°	2125.1±831.1°	2486.4±774.3°
TGFβ	100±6.48	389.02±37.03***	1416.45±228.28°°°	693.3±204.5	1064.3±405.7°°
Neutrophil apoptosis markers					
lactoferrin	100±10.58	83.09±23.29**	95.52±54.95	107.5±57.7	30.2±21.7
eNOS	100±5.95	116.38±11.70	279.52±38°°	255.56±21.34°°	311±40.8°°°
Extracellular matrix degradation markers					
RAGE	100±19.21	434.93±63.55***	813.96±123.17°	379.7±36.7	514.4±105.3
MMP2	100±6.48	4095.79***	10264±2497.28°°	2348.4±872.2	2779.4±815.9
TIMP2	100±7.09	954.77±118.80***	2021.2±140.55°°	1128.2±352.3	1336.8±307
Th17 cell marker					
IL-6	100±13.33	215.36±33.77**	82.89±49.37	49.55±8.04°	105.5±51.5
Treg cell marker					
Foxp3	100±13.41	1438.8±225.62	3188.2±1414.39°	766.4±544	332.1±103.7
Hepatic fatty acid Metabolism markers					
PPARα	100±4.11	71.28±6.11***	173.33±11.89°°°	214.9±26.3°°°	193.4±44.1°°
AR	100±4.67	118.78±11.85	211.39±13.88°	179.2±31.2	252.2±34.3°°
CD36	100±5.03	98±10.50	313.21±32.91°°°	179.5±33.4°	156.3±43
Lipid droplet Formation markers					
SNAP23	100±6.80	125.93±15.11	404.97±23.46°°	294.6±43.9°°	302±28.9°°°
VAMP4	100±7.26	95.99±9.45	194.84±9.01°°	151.5±28.7	187.7±13.05°°
Syntaxin 5	100±4.63	104.39±9.68	294.53±37.62°°	209.4±24.3°°	221.6±22.7°°°
Perilipin	100±16.38	255.29±48.62***	568.76±63.83°	761.4±250.8	1557.3±625.9
Insulin signaling markers					
IRS1	100±6.21	150.79±19.06**	268.60±33.07°	231.6±38.5	298.9±7°°°
SREBP1	100±9.30	202.77±33.01***	557.49±80.17°°	310.5±108.7	306.1±104.6
G6Pase	100±7.84	37.81±4.85***	259.07±58.26°°°	247.51±33.7°°°	184.4±61.1°°°
PEPCK	100±4.62	53.71±5.18***	157.71±3.68°°°	158.24±12.1°°°	160±35.5°°°



## Conclusions

In conclusion, **INT-767 treatment counteracts NASH in a rabbit model of HFD-induced MetS** by promoting insulin sensitivity, resolution of inflammation and fibrosis regression.

## References

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